

Assay of Cefetamet Pivoxil Hydrochloride in bulk and dosage forms by Visible Spectrophotometric methods

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Abstract : Simple, accurate and reproducible UV-Visible spectrophotometric methods were established for the assay of CPH (Cefetamet Pivoxil Hydrochloride) based on the formation complex products. Method A, Method B involves the redox/complex reactions of the CPH. The optical characteristics such as Beers law limits, molar absorptivity and Sandell's sensitivity for the methods (A-B) are given. Regression analysis using the method of least squares was made to evaluate the slope(b), intercept(a) and correlation coefficient (r) and standard error of estimation (Se) for each system. Determination of CPH in bulk form and in pharmaceutical formulations were also incorporated,

Key words: Determination, redox, complex, CPH

Introduction:

Cefetamet, [6R-[(6a,7β(Z))]]-7, [[(2-amino-4-

thiazolyl)(methoxyimino)acetyl]amino]-3-methyl-8-oxo-5-thia-1-azabicyclo-[4,2-

O]oct-2-ene-2-carboxylic acid (CPH), is an oral third-generation cephalosporin which is hydrolyzed to form the active agent cefetame[1-5]t. Cefetamet, because of its broad coverage of most gramnegative and gram-positive communityacquired pathogens, is one of the drugs of choice in the empiric therapy of respiratory and urinary communityacquired-infections[6,7] Literature survey serves only HPLC method[8-15] for analytical estimation of CPH; however, no spectroscopic studies for its estimation have been reported till date. Hence it was thought worthwhile to develop spectrophotometric method for the same. As the analytically useful functional groups in CPH have not been For Methods: fully exploited for designing suitable, visible spectrophotometric methods and

so still offer a scope to develop more visible spectrophotometric methods with better sensitivity, selectivity, precision and accuracy. The author has made some attempts in this direction and succeeded in developing thirteen visible spectrophotometric methods. All these methods have been extended to bulk and in its pharmaceutical formulation as well are described.

Experimental

i) Instruments used:

An Elico, UV-Visible digital spectrophotometer (SL - 159) with 1cm matched quartz cells were used for the spectral and absorbance measurements. An Elico LI-120 digital pH meter was used for pH measurements.

ii) Preparation of standard drug solutions:

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The stock solution (1mg/ml) of CPH was prepared by dissolving 100mg of it in 100ml 0.1M HCI. This solution was further diluted step wise with distilled water to obtain working standard solution of corresponding concentrations 100 μ gml⁻¹ [, M₁], 200 μ gml⁻¹[M₂],

iii) Proposed procedures:

After systematic and detailed study of the various parameters involved, as described under results and discussions the following procedures; [Methods Fe(III)/O-Phen (M₁); $Fe(III)/K_3Fe(CN)_6$ (M₂)] were recommended for the assay of CPH in bulk samples and pharmaceutical formulations.

a) For Bulk samples

Method - M₁

Aliquots (0.5-3.0mL, 100µg.mL-1) of standard CPH solution were transferred into a series of 10mL calibrated tubes and then solutions of 0.5mL (1.1 x 10-2M) of Fe (III), 2.0mL of (1.18 x 10-2M) ophenanthroline were added successively. The total volume in each tube was brought to 5.0mL with distilled water. The tubes were kept on a boiling water bath for 30min. The tubes were removed and cooled to room temperature. Two milliliters of (2.0 x 10-2M) o-phosphoric acid was added and volume in each tube was made up to the mark (10mL) with distilled water. The absorbance of the colored complex solution was measured after 5 min. At 530nm against a reagent blank prepared similarly. The content of the drug was computed from the appropriate calibration graph (Fig. 8).

Method $-M_2$

Into a series of 10.0mL calibrated tubes, aliquots of standard CPH solution

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(0.5 -2.5mL, 200µg.mL⁻¹) were transferred and 1.0mL of (3.32x10-3M) FeCl₃ solution was added . The tubes were stoppered immediately and shaken well for 5min. Then 0.5mL of 3.02 x 10^3 M Potassium ferricyanide solution was added into each tube and was closed with lids immediately. After 5min. 1.0mL of 1 N HCI was added and the final volume was added upto 10.0mL with distilled water. The absorbance of the solution in each tube was measured immediately at 710nm against a similar reagent blank. The amount of the drug was calculated from its calibration graph (Fig 4).

b) For pharmaceutical formulations:

An accurately weighed portion of tablet content equivalent to about 100 mg of CPH was transferred into a 100mL volumetric flask. Added about 80mL of warm isopropyl alcohol and shaken well for about 20min. The contents were diluted with isopropyl alcohol up to the mark and mixed thoroughly. The solution was filtered. The filtrate was evaporated to dryness. The residue was used for the preparation of formulation solutions for different methods as given under standard solutions preparations. These solutions were analyzed as under procedures described fro bulk solutions.

Results and Discussions:

i. Spectral Characteristics:

In order to ascertain the optimum wavelength of maximum absorption (λ_{max}) of the colored species formed in the above methods, specified amounts of CPH were taken and colors were developed separately by following the above procedures. The absorption spectra were scanned on a spectrophotometer in the wave length region of 340 to 900nm against similar reagent blank or distilled



water. The reagent blank absorption spectrum of each method was also recorded against distilled water. The results were graphically represented in Fig.1&2. The absorption curves of the

colored species in each method show characteristics absorption maxima where as the blank in each method has low or no absorption in this region.



Fig. 1: Absorption spectrum of CPH with Fe(III) - O-Phen (M_1)

Fig. 2: Absorption spectrum of CPH with $Fe(III) - K_8 Fe(CN)_8 (M_2)$



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Fig. 4: Beer's Law plot of CPH with $Fe(III) - K_3Fe(CN)_6 (M_2)$

0.7

0.6

0.5



ii. Optimum conditions fixation in procedures:

The optimum conditions for the color development of methods $(M_1, and M_2)$ were established by varying the parameters one at a time, keeping the others fixed and observing the effect produced on the absorbance of the colored species. The following experiments were conducted for this purpose and the conditions so obtained were incorporated in recommended procedures.

Method - M₁ [Fe (III)/o-phenanthroline]

In order to establish optimum conditions necessary for rapid and quantitative formation of the colored complex with maximum stability and sensitivity, the author has performed control experiments by varying one and fixing the other parameters, such as Absurbance 0.4 0.3 $Fe(III) = 1.80 \times 10^{-4} M$ 0.2 $K_3 Fe(CN)_6 = 1.51 \times 10^{-4} M$ $HCI = 1.00 \times 10^{-1} M$ 0.1 0 0.8 1.6 2.4 3.2 0 4 4.8 Drug concentration in $\mu g.mL^1$

effect of pH of the buffer solution, volume of buffer solution, volume of Fe (III) and o-phenanthroline solution, temperature, heating time, order of addition of reagents and nature of solvents for final dilution.

Method – M_2 [Fe(III) – K_0 Fe(CN)₀]

The optimum conditions in this method were fixed basing on the study of the effects of various parameters such as volumes of 3.32×10^{-3} M ferric chloride solution, 3.02×10^{-3} M potassium ferricyanide solution and 1N HCI, time and temperature necessary for complete color development, the stability and intensity of the colored species after final dilution were established by measuring absorbance's at 740 nm.

iii. Optical Characteristics:

In order to test whether the colored species formed in the above methods,

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adhere to Beer's law the absorbance's at appropriate wave lengths of a set of solutions containing varying amounts of CPH and specified amounts of reagents (as given in the recommended procedures for each method) were recorded against the corresponding reagent blanks. The Beer's law plots of these systems are recorded graphically (Figs.3 to 4) against the corresponding reagent blanks. Beer's law limits, molar absorptivity, Sandell's sensitivity and optimum photometric range (Table.1) for CPH in each method developed. With mentioned reagents were calculated. Least square regression analysis was carried out for getting the slope, intercept correlation and coefficient values. (Table1).

iv. Precision:

The precision of each proposal methods was ascertained from the absorbance values obtained by actual determination of six replicates of a fixed amount of CPH in total solution. The percent relative standard deviation and percent range of error (at 0.05 and 0.01 confidence limits) were calculated for the proposed methods (Table. 1).

v. Accuracy:

To determine the accuracy of each proposed method, different amounts of

bulk samples of CPH within the Beer's law limits were taken any analyzed by the proposed method. The results (percent error) are recorded in (Table1).

vi. Interference studies:

The effect of wide range of excipients and other active ingredients usually present in the formulations for the assay of CPH in methods (M_1 and M_2) under optimum conditions were investigated. The commonly used execipients and other active ingredients usually present in formulations did not interfere even if they were present in amount than they usually exist.

vii. Analysis of formulations:

Commercial formulations (tablets) containing CPH were successfully analyzed by the proposed methods. The values obtained by the proposed and reference methods for formulations were compared statistically with F and t tests and found not to different significantly. The results are summarized in (Table 2). Percent recoveries were determined by adding standard drug to preanalysed formulations. The results of the recovery experiments by the proposed methods are also listed in (Table 2).

_Table 1: Optical and regression characteristics, precision and accuracy of the proposed methods for CPH

| Parameter | M ₁ | M ₂ | |
|---|----------------|-------------------------|--|
| λ _{max} (nm) | 530 | 710 | |
| Bær's law limits (µg/mL) | 2.0-10.0 | 0.8-4.0 | |
| Detection limit (µg/mL) | 0.0353 | 0.1584 | |
| Molar absorptivity (1 mol ⁻¹ .cm ⁻¹) | 4.171 x 10⁵ | 8.939 x 10 ⁴ | |
| Sandell's sensitivity (µg.cm ⁻² /0.001 | 0.08104 | 0.08753 | |

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| absorbanœ unit) | | | | |
|---|--------------------------|---------------------------|--|--|
| Optimum photometric range (µg/mL) | 0.5-1.5 | 0.1-1.5 | | |
| Regression equation (Y=a+bc) | 7 435 x 10 ⁻¹ | | | |
| slope (b) | 1.400 × 10 | 1.686 x 10 ⁻¹ | | |
| Standard deviation on slope (S_b) | 1.325 x 10 ⁻³ | 5.396 x 10 ⁻³ | | |
| Intercept (a) | 2.0 x 10 ⁻³ | - 5.10 x 10 ⁻³ | | |
| Standard deviation on intercept (S _a) | 8.791 x 10 ⁻³ | 1.143 x 10 ⁻³ | | |
| Standard error on estimation (S_e) | 8.382 x 10 ⁻³ | 1.365 x 10 ⁻³ | | |
| Correlation coefficient (r) | 0.9995 | 0.9986 | | |
| Relative standard deviation (%)* | 4.2262 | 0.07161 | | |
| % Range of error (confidence limits) | | | | |
| 0.05 level * | 4.4350 | 0.07522 | | |
| 0.01 level | 6.9566 | 1.1785 | | |
| % error in Bulk samples ** | | | | |

* Average of six determinations considered

** Average of three determinations

Table 2: Assay of CPH in Pharmaceutical Formulations

| Formulations * | Amo unt | Amount found by proposed Methods** | | Reference method | Percentage recovery by proposed methods*** | | |
|-------------------|------------|---|--|--------------------------|---|-------------------------|--|
| | n (mg) | M ₁ | M ₂ | | M ₁ | M ₂ | |
| Tablet I | 250 | 249.23 <u>+</u> 0.7 ⁻ 1 F=1.828 t=1.555 | 249.36 <u>+</u> 0.5 4 F=3.160 t=1.431 | 249.98 <u>+</u> 0. 96 | 99.69 <u>+</u> 0.98 | 99.75 <u>+</u> 0.9 8 | |
| Tablet II | 500 | 499.48 <u>+</u> 0.5 7 F=2.719 t=1.009 | 499.15 <u>+</u> 0.7 0 F =1.803 t =1.626 | 499.52 <u>+</u> 0. 79 | 99.91 <u>+</u> 0.11 | 99.84 <u>+</u> 0.4 5 | |

* Tablets from four different pharmaceutical companies.

** Average <u>+</u> standard deviation of six determinations, the t-and F-test values refer to comparison of the proposed method with the reference method. Theoretical values at 95% confidence limit, F = 5.05, t = 2.262

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*** Recovery of 10mg added to the preanalysed pharmaceutical formulations (average of three determinations).

The proposed methods exploit the various functional groups in CPH molecule. Statistical analysis of the results shows that the proposed procedures have good precision and accuracy with good sensitivity and higher Results of the analysis of λ_{max}. pharmaceutical formulations reveal that the proposed methods are suitable for their analysis with virtually no interference of the usual additives present in pharmaceutical formulations.

The order of sensitivity among the proposed methods is: $M_1 > \dot{M_2}$.

Thus, the proposed methods are simple, sensitive or selective with reasonable precision and accuracy and constitute better alternatives to the reported ones in the assay of CPH in bulk form and pharmaceutical formulations

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